

A81

PCT

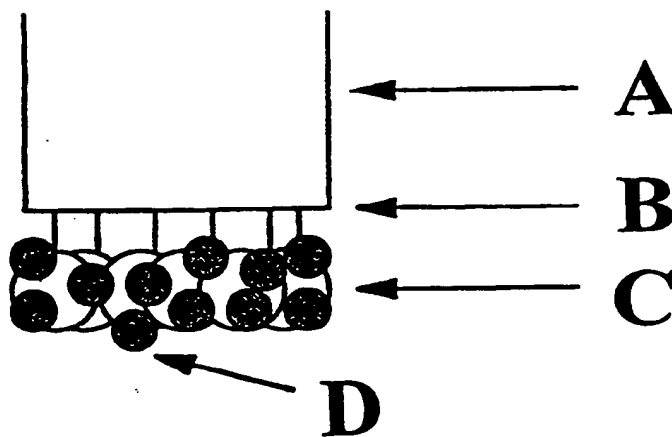
WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : G02B 6/00	A1	(11) International Publication Number: WO 99/17139 (43) International Publication Date: 8 April 1999 (08.04.99)
(21) International Application Number: PCT/US98/20081 (22) International Filing Date: 25 September 1998 (25.09.98) (30) Priority Data: 08/939,214 29 September 1997 (29.09.97) US (63) Related by Continuation (CON) or Continuation-in-Part (CIP) to Earlier Application US 08/939,214 (CIP) Filed on 9 September 1997 (09.09.97) (71) Applicant (for all designated States except US): THE REGENTS OF THE UNIVERSITY OF MICHIGAN [US/US]; 3003 South State Street, Ann Arbor, MI 48109-1280 (US). (72) Inventors; and (75) Inventors/Applicants (for US only): KOPELMAN, Raoul [US/US]; 1065 Heatherway, Ann Arbor, MI 48104 (US). CLARK, Heather [US/US]; 4519 Hunt Club Drive #2b, Ypsilanti, MI 48197 (US). BARKER, Susan [US/US]; 2573 Stone Road, Ann Arbor, MI 48105 (US).		(74) Agents: CARROLL, Peter, G. et al.; Medien & Carroll, LLP, Suite 2200, 220 Montgomery Street, San Francisco, CA 94104 (US). (81) Designated States: AL, AM, AT, AU, AZ, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i>

(54) Title: OPTICAL SENSORS FOR THE DETECTION OF NITRIC OXIDE**(57) Abstract**

Optical fiber sensors and fiberless optical sensors for measuring analytes, and in particular nitric oxide, are described utilizing metals, and more particularly, metal colloids. Proteins (or fragments thereof) with selective binding are immobilized on metal particles. The proteins may be dye-labeled for increased sensitivity.



FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AL	Albania	ES	Spain	LS	Lesotho	SI	Slovenia
AM	Armenia	FI	Finland	LT	Lithuania	SK	Slovakia
AT	Austria	FR	France	LU	Luxembourg	SN	Senegal
AU	Australia	GA	Gabon	LV	Latvia	SZ	Swaziland
AZ	Azerbaijan	GB	United Kingdom	MC	Monaco	TD	Chad
BA	Bosnia and Herzegovina	GE	Georgia	MD	Republic of Moldova	TG	Togo
BB	Barbados	GH	Ghana	MG	Madagascar	TJ	Tajikistan
BE	Belgium	GN	Guinea	MK	The former Yugoslav Republic of Macedonia	TM	Turkmenistan
BF	Burkina Faso	GR	Greece	ML	Mali	TR	Turkey
BG	Bulgaria	HU	Hungary	MN	Mongolia	TT	Trinidad and Tobago
BJ	Benin	IE	Ireland	MR	Mauritania	UA	Ukraine
BR	Brazil	IL	Israel	MW	Malawi	UG	Uganda
BY	Belarus	IS	Iceland	MX	Mexico	US	United States of America
CA	Canada	IT	Italy	NE	Niger	UZ	Uzbekistan
CF	Central African Republic	JP	Japan	NL	Netherlands	VN	Viet Nam
CG	Congo	KE	Kenya	NO	Norway	YU	Yugoslavia
CH	Switzerland	KG	Kyrgyzstan	NZ	New Zealand	ZW	Zimbabwe
CI	Côte d'Ivoire	KP	Democratic People's Republic of Korea	PL	Poland		
CM	Cameroon	KR	Republic of Korea	PT	Portugal		
CN	China	KZ	Kazakhstan	RO	Romania		
CU	Cuba	LC	Saint Lucia	RU	Russian Federation		
CZ	Czech Republic	LI	Liechtenstein	SD	Sudan		
DE	Germany	LK	Sri Lanka	SE	Sweden		
DK	Denmark	LR	Liberia	SG	Singapore		
EE	Estonia						

OPTICAL SENSORS FOR THE DETECTION OF NITRIC OXIDE

FIELD OF THE INVENTION

The invention relates generally to optical sensors, methods of sensor fabrication and uses of such sensors, and more particularly the use of such sensors for the
5 detection of nitric oxide.

BACKGROUND

Recent elucidation of the fact that nitric oxide plays many biological roles has spurred special interest in this molecule. For instance, nitric oxide is believed to play
10 a role in vasodilation. See Marletta *et al.*, "Unraveling the biological significance of nitric oxide" *Biofactors* 2:219 (1990). Nitric oxide also appears to inhibit platelet aggregation by elevating intraplatelet levels of cyclic GMP. See Diodati *et al.*, "Complexes of Nitric Oxide with Nucleophiles as Agents for the Controlled Biological Release of Nitric Oxide: Antiplatelet Effect" *Thrombosis and Haemostasis* 70:654
15 (1993)

More recently, nitric oxide is emerging as one of the main neurotransmitters in the central and peripheral nervous systems. See Snyder, "Janus faces of nitric oxide" *Nature* 364:577 (1993). It appears to play both neurotoxic roles, such as in AIDS dementia, and neuroprotective roles in degenerative problems such as Parkinson's and
20 Huntington's diseases.

Given the growing importance of the molecule, there have been a number of attempts to develop means to measure cellular levels of nitric oxide. For example, a fiber optic nitric oxide chemiluminescent sensor has been developed. See Zhou and Arnold, "Response Characteristics and Mathematical Modeling for a Nitric Oxide
25 Fiber-Optic Chemical Sensor" *Anal. Chem.* 68:1748 (1996). This sensor was constructed by holding a small amount of an internal reagent solution at the tip of a fiber-optic bundle with a piece of gas-permeable membrane. Nitric oxide diffuses across the membrane into this internal solution, where a chemiluminescent reaction

between nitric oxide, hydrogen peroxide, and luminol takes place. The drawbacks of this sensor include the following: 1) the response time (approximately 8-17 seconds) is longer than the time needed for nitric oxide in the solution to be converted to nitrite; 2) the detection of nitric oxide is complicated by interferences from dopamine, uric acid, ascorbic acid, and cysteine, 3) the sensor is relatively large in size (greater than 6 mm in diameter) and thus difficult to use for the measurement of cellular nitric oxide levels (and impossible for intracellular measurements); and 4) the sensor has relatively poor sensitivity, *i.e.* a relatively high limit of detection (approximately 1.3 mM of nitric oxide).

Sensors involving sol-gel technology have also been attempted. The process involves hydrolyzing an alkoxide of silicon to produce a sol, which then undergoes polycondensation to form a gel. Biomolecules are immobilized by being entrapped in the sol-gel. In one case, horse-heart cytochrome c was encapsulated in a sol-gel and absorbance-based spectral shifts were used to monitor the binding of nitric oxide. See Blyth *et al.*, "Sol-Gel Encapsulation of Metalloproteins for the Development of Optical Biosensors for Nitrogen Monoxide and Carbon Monoxide" *Analyst* 120:2725 (1995). Unfortunately, the sensor reaction is reported to have taken two hours to reverse, making dynamic measurements impossible.

What is needed is a sensor of relatively small size and good sensitivity that measures nitric oxide with little or no interference from other analytes in a short enough time period to permit dynamic measurements.

SUMMARY OF THE INVENTION

The invention relates generally to optical sensors, methods of sensor fabrication and uses of such sensors, and more particularly the use of such sensors for the detection of nitric oxide. The present invention contemplates both fiber-optic sensors and optical fiberless sensors comprising nitric oxide-binding compounds, such compounds permitting the specific binding of nitric oxide (e.g. non-covalent binding) with little or no interference from other analytes.

With regard to fiber-optic sensors, the present invention contemplates an optical fiber having a fiber tip, said tip comprising an immobilized nitric oxide-binding compound. It is not intended that the present invention be limited by the means by which the nitric oxide-binding compound is immobilized. In one embodiment, the tip
5 of the fiber is treated so as to have reactive groups and the nitric oxide-binding compound is covalently linked directly to the fiber via the reactive groups. In another embodiment, the tip has an inert coating (*i.e.* inert relative to nitric oxide) such as a metal layer (preferably, a non-linear layer and more preferably, spheres comprising metal) and the nitric oxide-binding compound is immobilized on the metal layer. In a
10 preferred embodiment, the tip is treated to create reactive groups (*e.g.* thiol groups), spheres of metal colloid are attached to the tip via the reactive groups, and the nitric oxide-binding compound is immobilized on the metal colloid spheres.

It is not intended that the present invention be limited to the nature or dimensions of the metal layer. A variety of metals and metal colloids are
15 contemplated, including but not limited to, colloids of gold, silver, tungsten, thoriosol, antimony pentoxide, carbon, red iron oxide, titanium dioxide and platinum (available commercially from Vector Laboratories, Inc., Burlingame, CA; Nanoprobes, Inc., Stony Brook, NY; and Polysciences, Inc., Warrington, PA). In a preferred
20 embodiment, the metal layer is a monolayer of spheres comprising gold colloid, said spheres attached to an end of a fiber as a substrate for spontaneous attachment of the nitric-oxide-binding compound. While not limited to particular dimensions, the size of the gold colloid does produce a marked difference in the fluorescence intensity measured. The present invention contemplates colloid sizes (and in particular gold
25 colloid sizes) ranging from very small, 2 nm, to very large, 250 nm (and more preferably, between 5 nm and 100 nm), said colloids immobilized on the end of a fiber to provide a base for protein attachment. While a precise understanding of the mechanism for this phenomenon is not necessary in order to practice the invention, it is surmised the intensity changes seen in the fluorescence emission are not a result of surface coverage, and availability of sites for protein adsorption, but instead a

quenching or enhancement by the gold itself. In general, the optimum fluorescence is achieved with particles sizes of approximately 100 nm.

It is also not intended that the present invention be limited by the nature of the particular nitric oxide-binding compound. In one embodiment, the nitric oxide-binding compound is a porphyrin group- or heme group-containing protein. In another
5 embodiment, the nitric oxide-binding compound is a heme-binding protein. Regardless of whether the protein is a heme-group-containing protein or a heme-binding protein, in one embodiment, the present invention contemplates that the protein (or peptide) is dye-labeled (e.g. with dyes which can be used for protein labeling that do not react to
10 nitric oxide, such as Oregon Green dyes). This has been found to increase the signal to noise ratio of the sensors of the present invention.

It is not intended that the present invention be limited to specific heme-group-containing proteins. The heme-group-containing proteins are limited only in the respect that they bind nitric oxide, and more preferably, they bind nitric oxide
15 specifically (i.e. they do not bind interfering substances). The preferred heme-group-containing protein is cytochrome c' (as distinct from cytochrome c). It is not intended that the present invention be limited to the source of cytochrome c'. Nonetheless, preferred sources include, but are not limited to, microorganisms, more preferably bacterial sources, and more particularly, purple phototropic bacteria, aerobic
20 nitrogen-fixing bacteria, and facultatively denitrifying bacteria, and still more particularly, sources such as *C. vinosum*, *R. purpureus*, and *R. gelatinosa*.

Insects have been shown to have both heme group-containing proteins that bind nitric oxide (M.C. Ribeiro *et al.*, "Reversible Binding of Nitric Oxide by a Salivary Heme Protein from a Bloodsucking Insect," *Science* 260:539 (1993); J.G. Valenzuela
25 *et al.*, "A Salivary Nitrophorin (Nitric-Oxide-Carrying Hemoprotein) In The Bedbug *Cimex lectularius*," *J. Exper. Biol.* 198:1519 (1995)], as well as heme-binding proteins [P.L. Oliveira *et al.*, "A Heme-binding Protein from Hemolymph and Oocytes of the Blood-sucking Insect, *Rhodnius prolixus*," *J. Biol. Chem.* 270:10897 (1995)]. The

present invention contemplates both groups of proteins as useful in the preparation of optical sensors.

It is not intended that the present invention be limited to specific heme-binding proteins. The heme-binding proteins are limited only in the respect that they bind
5 nitric oxide, and more preferably, they bind nitric oxide specifically (i.e. they do not bind interfering substances). The preferred heme-binding protein is the heme-binding protein isolated and characterized from both the hemolymph and oocytes of the blood-sucking insect, *Rhodnius prolixus*.

The invention also contemplates optical fiberless sensors capable of detecting
10 nitric oxide. The sensors of the present invention are: (1) small enough to enter a single mammalian cell relatively non-invasively, (2) fast and sensitive enough to catch even minor alterations in the concentration of nitric oxide and (3) mechanically stable enough to withstand the manipulation of the sensor to specific locations within the cell. Importantly, the fiberless sensors of the present invention are non-toxic and
15 permit the simultaneous monitoring of several cellular processes.

In one embodiment, the present invention contemplates fiberless optical sensors comprising a nitric oxide-binding compound. It is not intended that the present invention be limited by the precise composition of the fiberless sensors. The fiberless sensors of the present invention are either solid or semisolid particles ranging in size
20 between approximately 1 micrometer and 1 nanometer in diameter, and more preferably, between 5 nanometers and 250 nanometers. The ultimate small size is attained by fine grinding and filtering or by micro-emulsion techniques used to form mono-disperse colloidal particles (rather than nano-fabrication). In one embodiment, the sensor is selected from the group consisting of polymer fiberless sensors,
25 acrylamide fiberless sensors, sol-gel fiberless sensors and metal fiberless sensors.

In one embodiment, the polymer fiberless sensors of the present invention
comprise a nitric oxide-binding compound (such as a porphyrin) and a polymer. It is not intended that the present invention be limited to a particular polymer. In one embodiment, the polymer is selected from the group consisting of poly(vinyl chloride),
30 poly(vinyl chloride) carboxylated and poly(vinyl chloride-co-vinyl acetate-co-vinyl

alcohol). In a particular embodiment, the polymer fiberless sensors further comprise an additive and a plasticizer.

In one embodiment, the acrylamide fiberless sensors of the present invention comprise polyacrylamide and a nitric oxide-binding compound. In a preferred
5 embodiment, the acrylamide fiberless sensors further comprise N,N-methylenebi-
(acrylamide) and the mixture is polymerized to a gel.

In one embodiment, the sol-gel fiberless sensors of the present invention comprise a nitric oxide-binding compound entrapped in a matrix, such as a silica sol. Where the compound is a protein, stabilizers can be used. The gels are typically aged
10 before use.

In one embodiment, the metal fiberless sensors of the present invention comprise a nitric oxide-binding protein (or peptide) in combination with a metal selected from the group consisting of gold, silver, platinum and alloys thereof (e.g. a gold/silver alloy). In one embodiment, the protein (or peptide) is dye-labeled (e.g.
15 with Oregon green 514). Typically, such metal fiberless sensors are made by combining between 0.01% and 1% , and more preferably approximately 0.1% protein or peptide (by weight) in colloid solution. Spontaneous adsorption of the protein to the metal surface takes place within minutes.

It is not intended that the present invention be limited by the manner in which
20 the sensors of the present invention are introduced into cells. In one embodiment, a buffered suspension of fiberless sensors is injected into the sample cell with a commercially-available pico-injector. In another embodiment, the fiberless sensors of the present invention are shot into a cell with a commercially-available particle delivery system or "gene gun" (such gene guns were developed and are now routinely
25 used for inserting DNA into cells). In other embodiments, the fiberless sensors of the present invention are remotely steered into a cell by photon pressure or "laser tweezers". This uses an infra-red laser beam which traps the particles or magnetically, by remotely steering magnetic nanoparticle pebbles (commercially available) into a cell.

It is also not intended that the present invention be limited by the detecting means. In one embodiment, the fiberless sensors of the present invention are addressed by laser beams (rather than fibers), and their fluorescent signals are collected and analyzed by procedures identical to those used for the fiber-tip nanosensors. See
5 U.S. Patents Nos. 5,361,314 and 5,627,922 to Kopelman *et al*, hereby incorporated by reference.

DESCRIPTION OF THE DRAWINGS

Figure 1 schematically shows one embodiment of an optical fiber sensor of the present invention having a monolayer of metal spheres for attachment of a nitric
10 oxide-binding protein.

Figure 2 shows the heme group of one embodiment of a nitric oxide-binding protein, *i.e.* cytochrome c'.

Figure 3 shows one embodiment of an optical array useful for testing the sensors of the present invention.

15 Figure 4 is a graph showing the response of one embodiment of an optical fiber sensor of the present invention, said fiber having cytochrome c' of *C. vinosum* immobilized on gold colloid.

Figure 5 is a graph showing the response of one embodiment of an optical fiber sensor of the present invention, said fiber having cytochrome c' of *R. purpureus*
20 immobilized on gold colloid.

Figure 6 is a graph showing the response of one embodiment of an optical fiber sensor of the present invention, said fiber having cytochrome c' of *R. gelatinosa* immobilized on gold colloid.

25 Figure 7 shows the reversibility of one embodiment of the sensor of the present invention.

DEFINITIONS

To facilitate understanding of the invention, a number of terms are defined below.

The term "analyte" is intended to comprise any substance within a cell, including but not limited to nitric oxide and Na^+ , K^+ , Ca^{++} , Cl^- , H^+ , as well as oxygen and glucose.

The term "bacteria" refers to any bacterial species including eubacterial and archaeobacterial species.

The term "chemical reaction" means reactions involving chemical reactants, such as inorganic compounds.

The present invention contemplates "metal conjugates" such as conjugate comprising a nitric oxide-binding compound attached to a metal, including but not limited to, metal in the form of a metal colloid.

Colloids are solutions of dispersed or suspended particles. Where the particles comprise metal, the colloid is referred to as a "metal colloid".

The present invention contemplates "porphyrin group-containing proteins" and more preferably "heme group-containing proteins" capable of binding (and more preferably, binding selectively) nitric oxide. The porphyrins are named and classified on the basis of their side chain substituents, such as etioporphyrins, mesoporphyrins, uroporphyrins, coproporphyrins and protoporphyrins. The chelate complex of a porphyrin (e.g. protoporphyrin) with metal is contemplated by the present invention as useful for nitric oxide binding. The chelate complex with iron [e.g. Fe(II) , Fe(III)] is heme. Thus, the present invention specifically contemplates the use of ferrous and ferric hemoproteins to bind nitric oxide, and fragments thereof (preferably heme group containing fragments).

It is not intended that the present invention be limited by the type of porphyrin group-containing protein. In one embodiment, the present invention contemplates cyclases, including but not limited to, guanylate cyclase (which contains one protoporphyrin-IX type heme/heterodimer) and fragments thereof. *See generally* Stone and Marletta, "Soluble Guanylate Cyclase from Bovine Lung: Activation with Nitric

Oxide and Carbon Monoxide and Spectral Characterization of the Ferrous and Ferric States," *Biochemistry* 33:5636 (1994). In another embodiment, the present invention contemplates ferrous hemoproteins such as hemoglobin and myoglobin, or fragments thereof. In still another embodiment, the present invention contemplates cytochromes or portions of cytochromes as nitric oxide binding compounds. In a preferred
5 embodiment, the present invention contemplates cytochrome c' (as distinct from cytochrome c) and fragments thereof.

The present invention also contemplates compounds that bind porphyrins and heme. By the term "heme binding proteins" it is meant to indicate proteins that bind
10 heme groups. It is not intended that the present invention be limited to particular heme group-binding proteins. In one embodiment, the heme-binding compound is the 93,000 molecular weight, heme-binding protein from rabbit serum. See Tsutsui and Mueller, *J. Biol. Chem.* 257, 3925 (1982). In another embodiment, the heme-binding protein is selected from the group consisting of globin, serum albumin, and transferrin.
15 In yet another embodiment, the present invention contemplates the heme-binding protein isolated and characterized from both the hemolymph and oocytes of the blood-sucking insect, *Rhodnius prolixus*.

A compound is "immobilized" by attachment (whether covalent or non-covalent) or by encapsulation (e.g. in a gel such as a sol-gel).

20 In one embodiment, the present invention contemplates that the nitric oxide-binding compound is a protein (or peptide) and the protein (or peptide) is dye-labeled. There are three major classes of commonly used reagents to label amines: succinimidyl esters, isothiocyanates and sulfonyl chlorides. Dyes containing such groups are commercially available from Molecular Probes, Inc. (Eugene, OR) and include, but are
25 not limited to, such fluorophores as cascade blue, isosulfan blue, fluorescein, naphthofluorescein, malachite green and Oregon green).

"Initiating a reaction" means causing a reaction to take place. Reactions can be initiated by any means (e.g. heat, wavelengths of light, addition of a catalyst, etc.)

"Interfering substances" are those substances that interfere with the measurement of nitric oxide. Such substances include but are not limited to nitrite, dopamine, uric acid, ascorbic acid, and cysteine.

5 The term "microorganism" as used herein means an organism too small to be observed with the unaided eye and includes, but is not limited to bacteria, viruses, protozoans, fungi, and ciliates.

"Reactive groups" are those chemical groups that are capable of reacting with (and thereby capable of securing) compounds or substances to the fiber tip. Such groups include groups having a S, N or O functionality, including but not limited to
10 thiol groups, carboxylic acids and amino groups.

A "solvent" is a liquid substance capable of dissolving or dispersing one or more other substances. It is not intended that the present invention be limited by the nature of the solvent used.

DESCRIPTION OF THE INVENTION

15 The invention relates generally to optical sensors, methods of sensor fabrication and uses of such sensors, and more particularly the use of such sensors for the detection of nitric oxide. The present invention contemplates both fiber-optic sensors and optical fiberless sensors comprising nitric oxide binding compounds, such compounds permitting the specific binding of nitric oxide with little or no interference
20 from other analytes. This nitric oxide sensors of the present invention are small (100 μm and submicrometer), very selective and can be used for dynamic, real-time measurements.

A. Fiber-Optic Sensors And Their Fabrication

Micro-fiberoptic sensors (100-1000 nm) ~~are based on optical grade silica fibers~~
25 pulled to submicron size. The pulled fiber tips are much less fragile than those of the electrochemical microsensors, which are made from pulled micropipettes. Traditionally, a dye-polymer matrix is attached to the tip, which is very durable and smooth and runs tightly bound to the tip, even during penetration of biological tissues. The matrix on

the end of the fiber often includes several components, such as a chromoionophore, an ionophore, and appropriate ionic additives, all trapped inside a polymer layer, so that no chemicals are free to diffuse throughout the cell. The effects of toxicity of the dyes are thus minimized. Also, the probe can be carefully positioned in the cell, allowing
5 any specific area to be imaged or monitored.

The nitric oxide detecting sensors of the present invention, by contrast, has a metal monolayer attached to the tip. The fabrication involves treatment of the fiber so as to add reactive groups. Thereafter, the metal monolayer is attached, followed by immobilization of the nitric oxide-binding protein (see Figure 1, where the fiber (A),
10 the reactive groups (B), the metal monolayer (C) and the attached protein (D) are shown).

B. Optical Fiberless Sensors And Their Fabrication

The present invention contemplates nitric oxide detecting sensors that are fiberless sensors or Probes Encapsulated By BioListic Embedding (PEBBLEs). While
15 a variety of such fiberless sensors are contemplated, the preferred fiberless sensors of the present invention are those made with metal particles, and in particular, metal colloids.

The fiberless sensors are particularly suitable for chemical analysis in mammalian cells, by inserting the sensors into the cell, and monitoring remotely. The
20 sensor particles or beads can be dispersed in buffer solution and pico-injected into a cell. The particles can be monitored singly, in groups located at different positions, or several different kinds can be injected for simultaneous measurements of several distinct intracellular ion or small molecule concentrations.

The fiberless sensors of the present invention are prepared by coating metal
25 (e.g. Au) colloids with a nitric oxide binding compound (e.g. cytochrome c') or a dye-labeled nitric oxide-binding compound. For some cellular applications, a phospholipid layer is added to the fiberless sensors contemplated by the present invention.

C. Uses Of The Present Invention

The invention will be useful for, among other things, in the identification of cellular and subcellular responses which lead to disease. The fields in which the optical sensors of the present invention will find application are vast, and include basic
5 research, diagnosis, and treatment of disease. Direct benefits to humans and the environment include the development of new drugs, understanding the synergistic response to complex mixtures of pollutants, and prevention of developmental and degenerative disorders.

As noted above, nitric oxide is emerging as one of the main neurotransmitters
10 in the central and peripheral nervous systems. Accurate detection and measurement of nitric oxide in cells may be used diagnostically, given the role of nitric oxide in AIDS dementia, Parkinson's and Huntington's diseases.

1. Metal Colloids On Optical Fibers

While the nitric oxide detection features of the present invention have been
15 underscored, the present invention brings elements that can be used more generically if desired. Specifically, the use of metal colloids on optical fibers can be used to bind proteins other than nitric oxide binding proteins.

For example, an environmentally sensitive probe can be prepared using a metal layer (e.g. gold colloid) on the fiber tip. A dye can be attached to the end of a fiber
20 through a non-fluorescent protein (such as bovine serum albumin) and used as an indicator of the hydrophobicity of differing regions of a cell. All that has to be done is to use a dye which can easily be attached to a protein, and which is sensitive to changes in environment. Alternatively, any protein or peptide (such as a zinc finger peptide) which is sensitive to environmental changes or selectively binds certain
25 analytes (such as Zn^{2+}) can be used. Such proteins/peptides may be naturally fluorescent or labeled with fluorescent dyes. Attaching dyes to proteins can also be a useful method for measuring analytes in general.

Thus, the present invention contemplates fiber-optic sensors comprising an optical fiber having a fiber tip, said tip comprising an immobilized protein and a dye.

The protein can be any protein that is inert (*i.e.* inert relative to analyte sought to be detected). The protein can be immobilized using a metal layer (preferably, a non-linear layer and more preferably, spheres comprising metal). In one embodiment, the tip of the fiber is treated so as to have reactive groups and the spheres of metal colloid are attached to the tip via the reactive groups. In this latter embodiment, a fiber tip/reactive group/metal colloid/immobilized protein/selective dye complex is created.

2. Fiberless Sensors Comprising Metal Particles

As noted above, the nitric oxide detection features of the fiberless sensors of the present invention have been underscored. However, it is contemplated that fiberless sensors of the present invention can be used to measure any alteration in any endogenous analytes of any cell. The present invention specifically contemplates transcutaneous monitoring (*e.g.* ear, skin) as well as continuous flow monitoring of cells in culture, organotypic culture, organ slices, isolated perfused organs, organs *in situ*, and whole animal monitoring.

The present invention contemplates that the fiberless sensors of the present invention can be used as diagnostic tools for earlier intervention (*i.e.* earlier than currently available) and treatment of disease.

In one embodiment, the metal fiberless sensors of the present invention comprise a protein (or peptide) conjugated to a metal such as a metal colloid. The protein can be any protein that is inert (*i.e.* inert relative to analyte sought to be detected), and linked to a dye. Alternatively, any protein or peptide which is sensitive to environmental changes or selectively binds certain analytes (such as Zn^{2+}) can be used. Such proteins/peptides may be naturally fluorescent or labeled with fluorescent dyes. The protein can be immobilized using metal particles (preferably, a non-linear and more preferably, spheres comprising metal). In this latter embodiment, a metal colloid spherical particle/immobilized protein/selective dye complex is created.

3. Applications

Various fiber-optic tip sensors with fiberless ("pebble") sensors can be used. The fiber-optic nanotip fluorescent sensors are inserted into the cell by standard micropipette ("patch clamp") techniques. The chemical sensor nanotips and pebbles, with each tip or pebble specific to a given chemical analyte. Utilizing these optical nanosensors one can get a chemical video for a single analyte (say, calcium), analogous to a black and white movie, or of a list of chemical analyte (say, calcium, sodium, potassium, chloride, oxygen and pH), analogous to a color video or, alternatively, to six single color videos, each taken with a different narrow-band optical filter.

The cell is located under a microscope objective lens, and may be immobilized by a micropipette (standard technique). One or several fiber-optic nano-tips are inserted by fine manipulators. The fiber-optic tips may be multi-functional sensors (simultaneously measuring 2 or more analytes) and/or multiplexed fiber tips (simultaneously measuring 2 or more sensors). If sensor nanopebbles are inserted (via micropipette), they are addressed optically by laser beams, either directly or via optical fibers.

Elimination of cross-talk among the different nanosensors is based on a combination of spatial, temporal and spectral resolution. Different fiber-optic tip sensors are excited by different lasers or laser wavelengths. Image focusing on an intensified CCD (requested item) separates spatially the fluorescence from different nano-sensors. Spectral control of the excitation wavelength and spectral selection of the fluorescence wavelength separates the various signals.

EXPERIMENTAL

The following examples serve to illustrate certain preferred embodiments and aspects of the present invention and are not to be construed as limiting the scope thereof. In the experimental disclosure which follows, the following abbreviations
5 apply: TEA (triethylamine; EDTA (ethylenediaminetetraacetic acid); IBMX (isobutylmethylxanthine); DTT (dithiothreitol; GTP (guanosine 5'-triphosphate); SNP (sodium nitroprusside).

EXAMPLE 1

In this example, sensors were prepared by silanizing a freshly cleaved
10 multimode fiber by immersing the distal end for two hours in 3-(mercaptopropyl) trimethoxysilane, thereby modifying the fiber with reactive groups. The end was then rinsed copiously with methanol, then triply distilled water. The silanized fiber was placed in colloidal gold (used as received from the manufacturer) for three hours, then rinsed with water. The sensor can be stored at this point in water or air.

15 Two protein solutions were prepared. The first solution was prepared by dissolving 0.1% r-phycoerythrin (Molecular Probes, Eugene, OR) in phosphate buffer, pH 6.0. The fiber was then immersed in the protein solution for one hour, rinsed with buffer, then used.

A second protein solution was prepared by dissolving 0.1% bovine serum
20 albumin (BSA) in phosphate buffer, pH 6.0. The fiber was then immersed in the protein solution for one hour, rinsed with buffer, then used. After BSA attachment, the fiber tip was placed in Texas Red-X (Molecular Probes, Eugene, OR) solution for one hour. The dye solution was prepared by dissolving 1 mg Texas Red-X in dimethyl sulfoxide (DMSO) then adding 1 M sodium bicarbonate solution (pH 8.3).

25 The sensor was rinsed with water before use.

The complete optical path for the fiber optic sensors included: Ion Laser Technology (Salt Lake City, UT) argon ion laser; 514.5 nm laser band-pass filter (Newport Corp. Irvine, CA); Uniblitz shutter controller (Rochester, NY); fiber coupler (Newport Corp. Irvine, CA); Olympus inverted fluorescence microscope, IMT-II (Lake

Success, NY); Nikon 50 mm f/1.8 camera lenses; Acton 150 mm spectrograph (Acton, MA); and, a Princeton Instruments 1024x256 LN2 cooled CCD array (Trenton, NJ).

The results showed that, based on representative spectra of both the R-phycoerythrin and the Texas red labeled- bovine serum albumin, the proteins were
5 immobilized onto the end of an optical fiber (data not shown). More importantly, although there is only a monolayer of fluorescent material on the surface of the optical fiber, a strong signal was measured (data not shown). Interestingly, the results indicate that the size of the gold colloid used in the sensor development plays a significant role in the amount of fluorescence signal obtained, with 100 nm particles
10 showing what appear to be the best results (data not shown).

EXAMPLE 2

In this example, pulled sensors are described. Sub-micron sized fiber optic sensor were prepared by pulling multimode fiber in a home-built puller consisting of a modified pipette-puller heated with a CO₂ laser. The pulled tips were then silanized,
15 coated with aluminum in a home-built evaporator and prepared with gold colloid and protein as above.

EXAMPLE 3

This example describes the first sensor to incorporate cytochrome c'. The heme group of this nitric oxide-binding protein is shown in Figure 2. Cytochrome c'
20 was chosen as the chemical recognition element because it exhibits spectral changes upon binding nitric oxide and is highly selective. While a precise understanding of the mechanisms involved is not necessary for the practice of the invention, it is believed that, as the sixth ligand site of cytochrome c' is buried within the protein, it is usually accessible only to carbon monoxide and nitric oxide. However, cytochrome c'
25 undergoes autooxidation to Fe(III), which binds nitric oxide, but not carbon monoxide.

To prepare the sensor, 100 μ m core diameter multimode fibers (General Fiber Optics, Cedar Grove, NJ) or multimode fibers were pulled to a submicrometer tip diameter and were silanized for 2 hrs in neat 3-mercaptopropyl-trimethoxy silane

(Gelest, Inc., Tullytown, PA) in a well-ventilated fume hood. The fibers were rinsed with methanol and water and placed in 50 nm gold colloid (Vector Labs, Inc., Burlingame, CA) for 3 hrs at 0 °C to form a self-assembled monolayer of gold colloids. The gold-coated fibers were immersed overnight in either a *Chromatium vinosum*, *Rhodocyclus purpureus*, or *Rhodocyclus gelatinosus* cytochrome c' solution prepared with pH 7.4 phosphate buffer at 15°C to allow the cytochrome c' to adsorb to the surface of the gold via the protein amine groups. The fiber was then rinsed in buffer to remove any residual protein. Dye-labeled sensors were prepared with 4 - carboxy - 3,5,6 trifluoro - 2',7',- difluorofluorescein, succinimidyl ester (Oregon Green™ 514 carboxylic acid, succinimidyl ester) purchased from Molecular Probes (Eugene, OR). A 16 mM dye solution in DMSO was diluted to 1 M with 0.1 M NaHCO₃. The cytochrome c' sensors were soaked for 1 hr in the dye solution.

The gold colloid-containing sensors that were prepared with either *C. vinosum*, *R. purpureus*, or *R. gelatinosa* cytochromes c' were tested. One embodiment of the optical array useful for testing is shown schematically in Figure 3). All three cytochromes c'-containing sensors were found to have linear responses up to 1mM nitric oxide (Figures 4-6). Above this concentration is a second linear region with a slope which is three times that below 1 mM, as shown in Figure 7. The steeper slope found at higher concentrations may be due to spectroscopic changes upon binding a second nitric oxide molecule per monomer, or may be caused by each cytochrome c' dimer binding more than two nitric oxide molecules. In view of their structural similarities, it is not surprising that all three cytochromes c' have comparable slopes. The *C. vinosum* (Figure 7, where squares indicate increasing nitric oxide concentration and circles indicate decreasing nitric oxide concentration) and *R. purpureus* cytochromes c' are readily reversible. The *R. gelatinosa* cytochrome c' does not respond reversibly, but the reasons for this difference have not yet been determined. The present limit of detection is approximately 20 µM nitric oxide. While the reaction of *C. vinosum* cytochrome c' has been reported to be slow when measured in solution

via absorbance, the results using the sensors of the present invention show that the sensor's response time is less than 2 seconds.

The *C. vinosum*, *R. purpureus*, and *R. gelatinosa* cytochromes c' sensors were tested for selectivity against common interfering chemicals. 1M nitrate and nitrite, oxygen, nitrogen, and 1.5×10^{-4} M ascorbic acid, in phosphate buffer solution, were found to have no effect on the sensor response (data not shown). Low μ M riboflavin was found to interfere with dye-labeled sensors due to its fluorescence band in the 530 nm region, overlapping with that of the labeling dye. The addition of 4 μ M riboflavin was equivalent to a 30 μ M decrease in nitric oxide concentration. However, riboflavin does not affect the cytochrome c' fluorescence. Dopamine at high levels also interfered with the detection of nitric oxide. In solution, dopamine was found to cause a blue shift in the protein fluorescence. For sensors prepared without the labeling dye, the addition of 140 mM dopamine was, on average for the three cytochromes c', equivalent to the addition of 0.1 mM nitric oxide. For the dye-labeled sensors, 0.25 mM dopamine was, on average, equivalent to the addition of 0.1 mM nitric oxide. Sensors made with the cytochromes c' alone (without dye labeling) were not affected by pH between pH 3-10. The dye fluorescence is sensitive to pH below 7. In solution, hydrogen peroxide caused an attenuation of the cytochrome c' fluorescence, but no spectral shift. Dye-labeled sensors exhibited an average signal increase of 4% in 1M hydrogen peroxide, equivalent to the removal of 0.2 mM nitric oxide. However, the concentration of oxygen radicals needed to interfere with the sensor response is higher than that found in many biological systems.

EXAMPLE 4

In this example, a heme-binding protein is prepared and attached to a metal monolayer of a fiber tip. The heme-binding protein prepared is the protein that been isolated and characterized from both the hemolymph and oocytes of the blood-sucking insect, *Rhodnius prolixus*. See generally P.L. Oliveira *et al.*, "A Heme-binding Protein from Hemolymph and Oocytes of the Blood-sucking Insect, *Rhodnius prolixus*," *J. Biol. Chem.* 270:10897 (1995). The *Rhodnius* heme-binding protein (RHBP) is

composed of a single 15-kDa polypeptide chain coiled in a highly α -helical structure which binds non-covalently one heme/polypeptide chain. This RHBP is not produced by limited degradation of hemoglobin from the vertebrate host, since specific polyclonal antibodies against it do not cross-react with rabbit hemoglobin, and since it
5 differs from hemoglobin in having a distinct amino-acid composition and NH_2 -terminal sequence. The spectrum of the dithionite-reduced protein has peaks at 426, 530, and 559 nm and resembles that of a *b*-type cytochrome.

RHBP from hemolymph is not saturated with heme and promptly binds heme added to the solution. The oocyte protein, on the other hand, is fully saturated and is
10 not capable of binding additional heme.

For preparation of the protein, insects are taken from a colony of *R. prolixus* maintained at 28 °C and 70% relative humidity. Normal mated females are fed on rabbit blood at 2-week intervals. Four to 6 days after a meal, hemolymph is collected in the presence of phenylthiourea (30-130 $\mu\text{g}/\text{ml}$), 5 mM EDTA, and a mixture of
15 protease inhibitors prepared in 0.15 M NaCl, with final concentrations of 0.05 mg/ml of soybean trypsin inhibitor, leupeptin, lima bean trypsin inhibitor and antipain, and 1 mM benzamidine. On the same day, chorionated oocytes are dissected and washed with ice-cold 0.15 M NaCl in order to remove ovarian debris prior to homogenization. Oocytes are homogenized in a Potter-Elvehjem homogenizer in the presence of the
20 same mixture of protease inhibitors, buffered with 20 mM Tris-HCl, pH 7.0, (approximately 500 oocytes to 1 ml), and centrifuged at room temperature in a microcentrifuge at 11,000 x g for 5 min. The floating lipids and the pellet are discarded, and the clear supernatant is used as the crude oocyte extract for protein purification.

25 Solid ammonium sulfate is added to bring the oocyte extract to 45% saturation, and the suspension is gently stirred for 20 min at 4 °C. After centrifugation at 11,000 x g for 10 min, the precipitate is discarded, and the supernatant is brought to 60% saturation. This new precipitate is then washed twice with a 60% saturated ammonium sulfate solution and then back-extracted by resuspending in a 45% saturated solution

and centrifuging. The pellet is discarded and the supernatant is dialyzed against 0.15 M NaCl, 10 mM Tris-HCl, pH 7.0, and applied to a column of Sephadex G-200 (2.5 x 55 cm) equilibrated with the same solution. Protein content of fractions is measured by the absorbance at 280 nm. The colored fractions containing RHBP are pooled,
5 dialyzed against deionized water, and lyophilized.

Hemolymph (approximately 3 ml) is diluted to 5 ml with phosphate-buffered saline (0.15 M NaCl, 0.1 M sodium phosphate, pH 7.0) and 1.25 g of KBr is added. The solution is centrifuged at 80,000 x g for 20 h at 4 °C. The fractions at the bottom of the tube are collected and dialyzed against deionized water until an abundant
10 precipitate forms. The solutions is then centrifuged at 11,000 x g for 10 min at 4 °C. The supernatant is brought to 10 mM with Tris base and applied to a column (1.5 x 18 cm) of DEAE-Toyopearl, equilibrated with 10 mM Tris-HCl, pH 8.4. The column is first washed with 20 ml of the same buffer and then eluted with and NaCl gradient (0-100 mM). The fractions containing RHBP are pooled and applied to a Sephadex G-75
15 column (1.5 x 80 cm) equilibrated with 0.15 M NaCl, 10 mM Tris-HCl, 7.0. Fractions containing the RHBP are pooled, dialyzed against deionized water, and lyophilized.

Proteins from both sources are monomeric, as indicated by an apparent molecular weight of between approximately 12,000 and 15,000. The purified protein (0.01%) is added to metal colloid as described above for the other proteins.

20

EXAMPLE 5

In this example, a nitric-oxide-carrying hemoprotein is prepared from a salivary gland homogenate of the bedbug *Cimex Lectularius*. The insect colonies are maintained at 27 C and 65% humidity. Insects are fed every 10 days by exposing them to the shaved abdomen of an anesthetized rabbit. Salivary glands of insects at 8-
25 10 days after feeding are dissected and stored in buffers or used immediately. In any event, they are disrupted with a dissection needle and the homogenate centrifuged in the cold at 14,000 RPM for 5 minutes. The supernatants are recovered and used for binding to metal colloids as described above.

The salivary gland homogenates have a nitrosyl-hemoprotein that releases nitric oxide in a pH-dependent manner. The fraction containing the NO-carrying hemoprotein, when separated by HPLC, causes vasodilation of a precontracted rabbit aortic strip.

5

EXAMPLE 6

This example describes the preparation of soluble guanylate cyclase ("sGC"). See generally Stone and Marletta, "Soluble Guanylate Cyclase from Bovine Lung: Activation with Nitric Oxide and Carbon Monoxide and Spectral Characterization of the Ferrous and Ferric States," *Biochemistry* 33:5636 (1994). sGC can be purified from bovine lung. Steps should be performed at 4°C. During the purification, assays for sGC can be carried out in 50 mM TEA, pH 7.4, with 2 mM DTT, 5mM MnCl₂, 100 µM GTP, 1mM IBMX, 5mM phosphocreatine, and 152 units/L creatine kinase, with or without 100 µM sodium nitroprusside (SNP) in a total volume of 100 µL at 37°C. The assays are quenched with 400 µL of 125 mM Zn(CH₃COO)₂ and 500 µL of 125 mM Na₂CO₃. The amount of cGMP generated can then be quantiated via radioimmunoassay (Amersham). After each column, only active fractions which can be activated at least 10-fold with SNP are carried on to the subsequent step.

Fresh bovine lung (1100g) is homogenized with a food processor in 1100 mL of homogenizing buffer (25 mM TEA, 5mM DTT, 1mM PMSF, 1 mM EDTA, 0.2 mM benzamidine, 1µM pepstatin A, 1µM leupeptin, pH 7.8). The homogenate is then centrifuged at 100000g for 60 min. The resulting supernatant is added to to 450 mL of DEAE-Sepharose CL-6B and stirred slowly with an overhead stirrer for 45 min. The buffer is removed by vacuum filtration, and the resin washed four times by stirring slowly for 15 min with 1200 mL of homogenizing buffer. The resin is then poured into a 2.5 x 100 cm column and packed by washing the column with homogenizing buffer for 1 h at 80 mL/h. sGC is then eluted with 1 L of a 0.0-0.4 M NaCl gradient at 80 mL/h. Active fractions are pooled (140 mL) and dialyzed for 8 h against 4 L of 25 mM TEA, 5 mM DTT, 0.1 mM PMSF; 0.2 Mm benzamidine, 1 µM pepstatin A, 1µM leupeptin, ph 7.4.

The sample is then brought to 5mM MnCl_2 and applied μ 25 mL/h to a 75-mL column of ATP-agarose (11-atom spacer attached through C-8). The column is washed at 25 mL/h with 150 mL of buffer A (25 mM TEA, mM DTT; 1 mM PMSF, 0.2 mM benzimidazole, 5 mM MnCl_2 , pH 7.4). The column is then washed at 50 mL/h with 200 mL of buffer A containing 10 mM NaNO_3 , 10 mM creatine, and 5mM ADP followed by 300 mL of buffer A. sGC is then eluted at 50mL/h with a 2-L gradient running from 5 mM MnCl_2 to 1 mM EDTA. Active fractions are pooled (200 mL), concentrated by ultrafiltration to 30mL on a YM-30 membrane (Amicon), and brought to 5 mM MnCl_2 .

The sample is then applied at 5 mL/h to a 3.0-mL column of GTP-agarose (11-atom spacer attached through ribose hydroxyls, ICN). The column is washed at 5 mL/h with 10-mL of buffer A, and then sGC is eluted with a 100-mL gradient running from 0.0 to 0.4 M NaCl in buffer A. Active fractions are pooled, concentrated by ultrafiltration to 8 mL on an Omega Series membrane (30 kDA MWCO, Filtron), and applied to a 2.5 x 95 cm column of Ultrogel AcA-34 (IBF) at 20 mL/h. The column is then washed with 400 mL of 25 mM TEA, 5mM DTT pH 7.8 at 20mL/h. Active fractions are pooled (20mL), provided with additional DTT (5 mM final), brought to 1.0 M NaCl, and concentrated by ultrafiltration to 1.0 mL on an Omega Series membrane. The sample is brought to 50% glycerol and stored as aliquots under nitrogen at -70°C .

The above-described purification scheme results in the isolation of approximately 0.5 mg of sGC from 1100 g of bovine lung. Following the procedure, two bands should be visible on reducing SDS-PAGE of approximately equal intensity with apparent masses of 69 and 78 kDA. The protein prepared as described above can then be immobilized on metal colloids.

EXAMPLE 7

This example describes the preparation of macro-sensors (sensors made on larger substrates, *i.e.* larger than fiber optic sensor tips). The sensor was made on glass microscope slides cleaned by immersion in piranha solution (10:1 sulfuric acid
5 :hydrogen peroxide), copious rinsing in triply distilled water and oven drying. The glass was then silanized in a 10% solution of 3-(mercaptopropyl) trimethoxysilane in methanol for 24 hours. The glass was rinsed with methanol, then triply distilled water. The silanized glass was placed in colloidal gold (used as received from the manufacturer) for 24 hours, then rinsed with water. For comparison of sizes of gold
10 colloids, six different gold sizes were used: 5, 10, 20, 50, 100 and 250 nm.

From the above, it should be clear that the methods, devices and compositions of the present invention permit for the design of sensors that have excellent selectivity, reversibility, fast response time, and can be reduced to submicrometer scale for use in single-cell or other *in vitro* applications.

CLAIMS

1. An optical fiber comprising a fiber tip, said tip comprising an immobilized nitric oxide-binding compound.
2. The optical fiber of Claim 1, wherein said fiber tip is treated so as to have reactive groups and said nitric oxide-binding compound is covalently linked directly to said fiber via said reactive groups.
3. The optical fiber of Claim 1, wherein said tip is coated with an inert coating and said nitric oxide-binding compound is immobilized on said coating.
4. The optical fiber of Claim 3, wherein said inert coating comprises a metal layer.
5. The optical fiber of Claim 4, wherein said metal layer is a monolayer.
6. The optical fiber of Claim 5, wherein said metal monolayer is non-linear.
7. The optical fiber of Claim 6, wherein said metal monolayer comprises spherical metal particles.
8. The optical fiber of Claim 7, wherein said metal particles comprise metal selected from the group consisting of gold, silver, tungsten, thoriasol, antimony pentoxide, carbon, red iron oxide, titanium dioxide and platinum.
9. The optical fiber of Claim 1, wherein said nitric oxide-binding compound is a protein selected from the group consisting of porphyrin group- and heme group-containing proteins, and fragments thereof.

10. The optical fiber of Claim 9, wherein said nitric oxide-binding compound is the heme group-containing protein cytochrome c'.
11. The optical fiber of Claim 10, wherein said cytochrome c' is purified cytochrome c' from a bacteria.
- 5 12. The optical fiber of Claim 11, wherein said bacteria is selected from the group consisting of purple phototropic bacteria, aerobic nitrogen-fixing bacteria, and facultatively denitrifying bacteria.
13. The optical fiber of Claim 12, wherein said bacteria is selected from the group consisting of *C. vinosum*, *R. purpureus*, and *R. gelatinosa*.
- 10 14. The optical fiber of Claim 9, wherein said nitric oxide-binding compound is the heme group-containing protein guanylate cyclase.
- 15 15. The optical fiber of Claim 9, wherein said guanylate cyclase is bovine lung guanylate cyclase.
16. The optical fiber of Claim 1, wherein said nitric oxide-binding compound is a heme-binding protein.
17. The optical fiber of Claim 1, wherein said heme-binding protein is dye-labeled.
18. The optical fiber of Claim 16, wherein said heme-binding protein is an insect heme-binding protein.
- 20 19. The optical fiber of Claim 18, wherein said heme-binding protein is the heme-binding protein of *Rhodnius prolixus*.

20. The optical fiber of Claim 1, wherein said nitric oxide-binding compound is a protein selected from the group consisting of dye-labeled, porphyrin group- and dye-labeled, heme group-containing proteins, and fragments thereof.
21. An optical fiber comprising a fiber tip, said tip comprising an metal layer and
5 a protein or fragment thereof immobilized on said metal layer.
22. The optical fiber of Claim 21, wherein said metal layer is a monolayer.
23. The optical fiber of Claim 22, wherein said metal monolayer is non-linear.
24. The optical fiber of Claim 23, wherein said metal monolayer comprises spherical metal particles.
- 10 25. The optical fiber of Claim 24, wherein said metal particles comprise metal selected from the group consisting of gold, silver, tungsten, thoriasol, antimony pentoxide, carbon, red iron oxide, titanium dioxide and platinum.
26. The optical fiber of Claim 21, wherein said protein or fragment thereof is a nitric oxide-binding compound.
- 15 27. The optical fiber of Claim 26, wherein nitric oxide-binding compound is selected from the group consisting of porphyrin group- and heme group-containing proteins, and fragments thereof.
28. The optical fiber of Claim 27, wherein said nitric oxide-binding compound is the heme group-containing protein cytochrome c'.

29. The optical fiber of Claim 28, wherein said cytochrome c' is purified cytochrome c' from a bacteria.
30. The optical fiber of Claim 29, wherein said bacteria is selected from the group consisting of purple phototropic bacteria, aerobic nitrogen-fixing bacteria, and facultatively denitrifying bacteria.
31. The optical fiber of Claim 30, wherein said bacteria is selected from the group consisting of *C. vinosum*, *R. purpureus*, and *R. gelatinosa*.
32. The optical fiber of Claim 26, wherein said nitric oxide-binding compound is the heme group-containing protein guanylate cyclase.
33. The optical fiber of Claim 32, wherein said guanylate cyclase is bovine lung guanylate cyclase.
34. The optical fiber of Claim 26, wherein said nitric oxide-binding compound is a heme-binding protein.
35. The optical fiber of Claim 34, wherein said heme-binding protein is dye-labeled.
36. The optical fiber of Claim 34, wherein said heme-binding protein is an insect heme-binding protein.
37. The optical fiber of Claim 36, wherein said heme-binding protein is the heme-binding protein of *Rhodnius prolixus*.

38. The optical fiber of Claim 26, wherein said nitric oxide-binding compound is a protein selected from the group consisting of dye-labeled, porphyrin group- and dye-labeled, heme group-containing proteins, and fragments thereof.

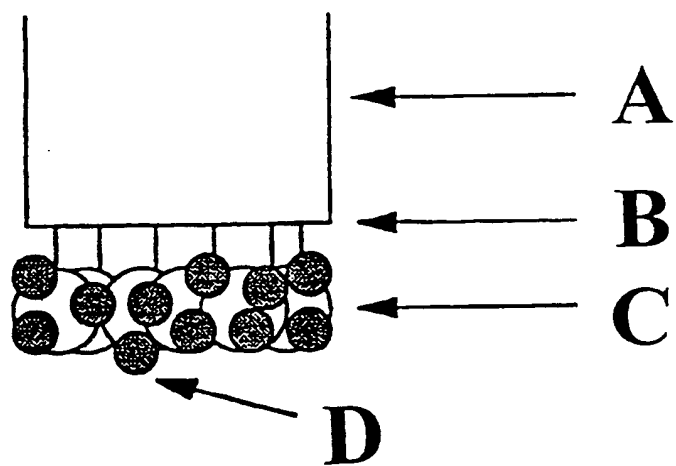


FIGURE 1

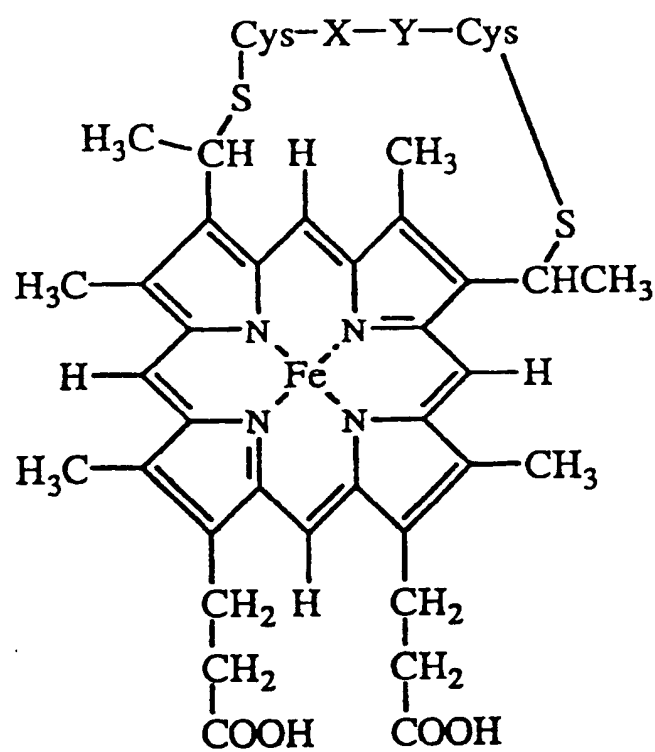


FIGURE 2

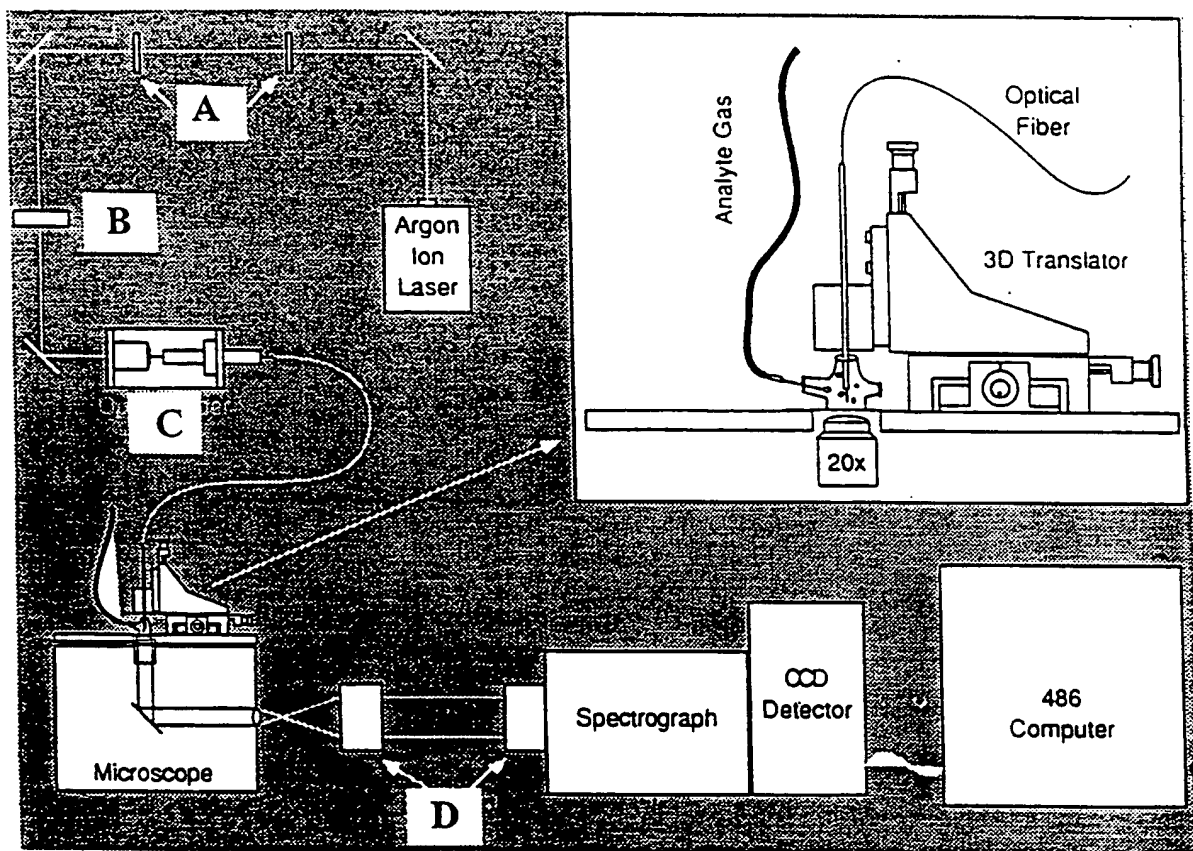


FIGURE 3

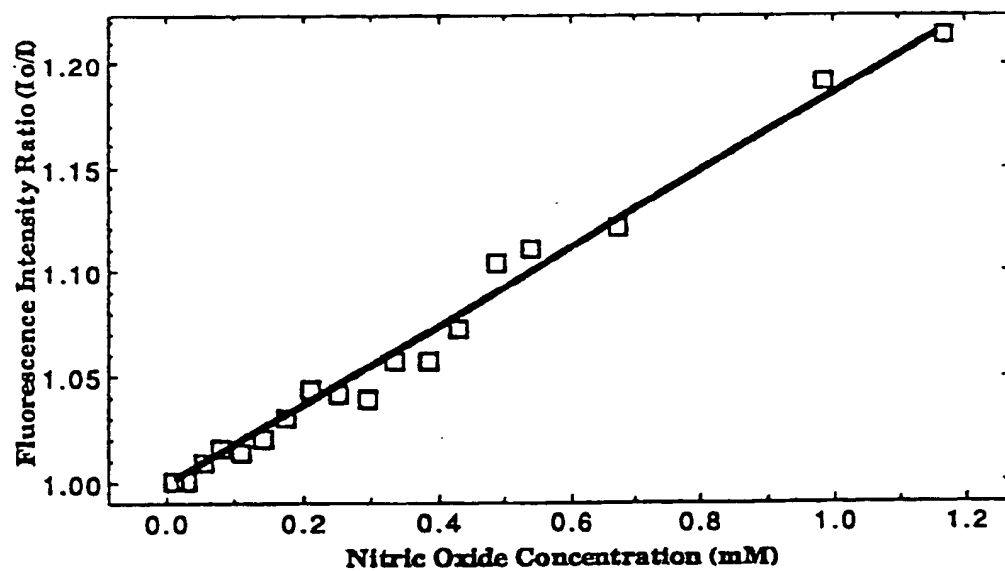


FIGURE 4

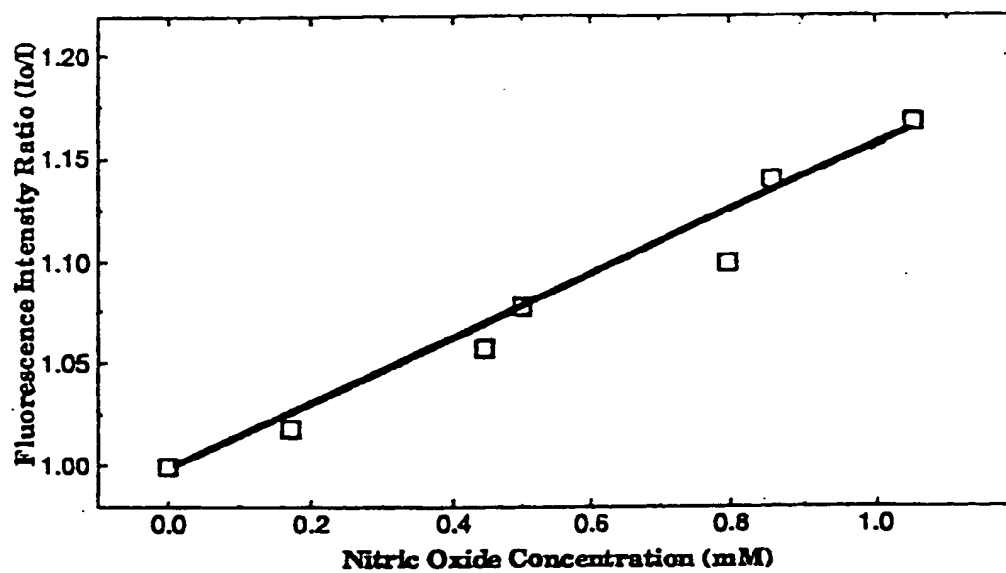


FIGURE 5

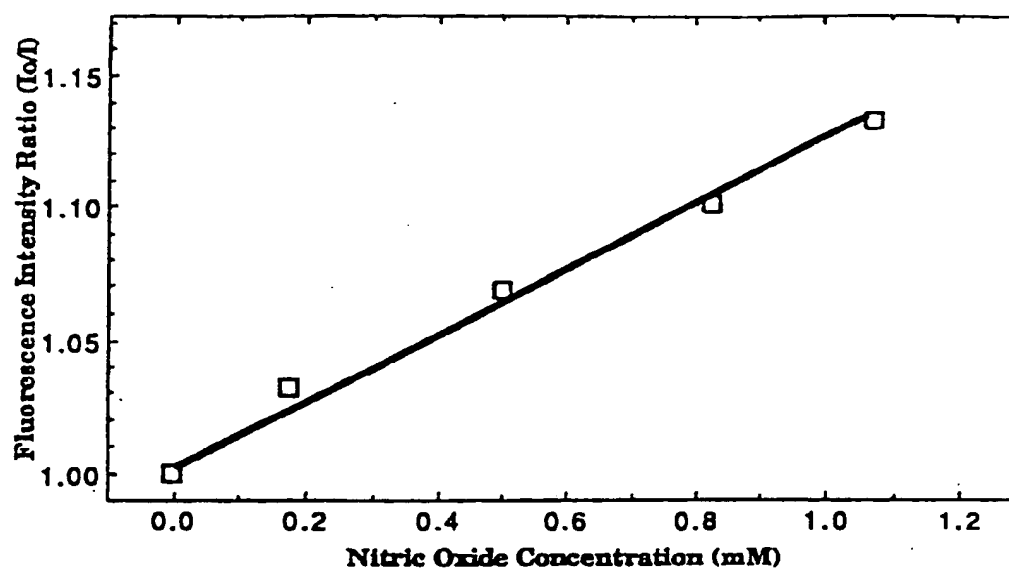
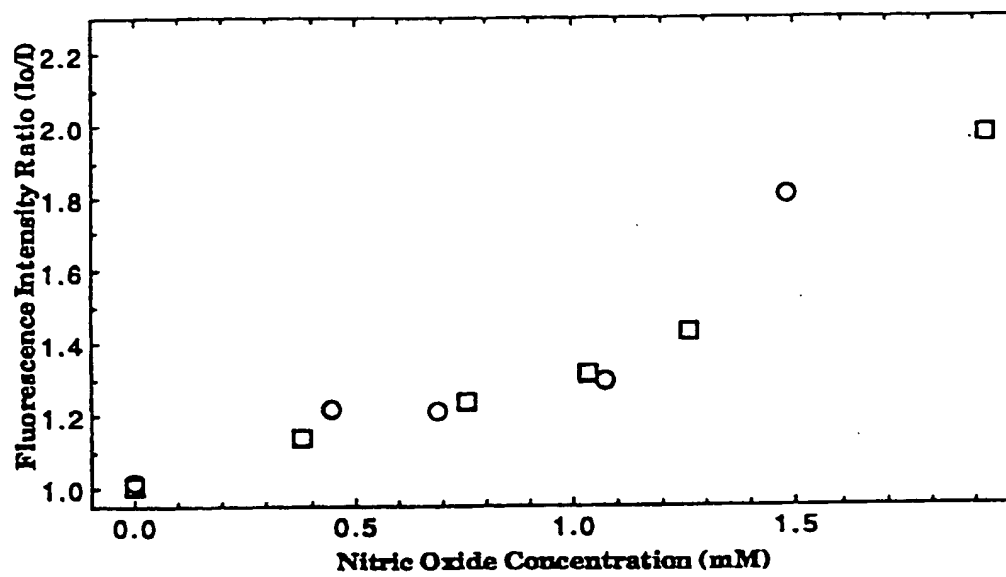


FIGURE 6

**FIGURE 7**

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US98/20081

A. CLASSIFICATION OF SUBJECT MATTER IPC(6) : G02B 6/00 US CL : 385/12, 117, 118, 31, 43 According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) U.S. : 385/12, 117, 118, 31, 43 385/14, 38, 127, 128, 141, 147 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) Please See Extra Sheet.		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 5,361,314 A (Kopelman et al.) 01 November 1994 (01-11-94), see figure 4	1-4 and 21
Y	US 5,627,922 A (Kopelman et al.) 06 May 1997 (06-05-97) see figure 4.	1-4 and 21
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.		
* *A* *E* *L* *O* *P*	Special categories of cited documents: document defining the general state of the art which is not considered to be of particular relevance earlier document published on or after the international filing date document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) document referring to an oral disclosure, use, exhibition or other means document published prior to the international filing date but later than the priority date claimed	*T* *X* *Y* *A* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art document member of the same patent family
Date of the actual completion of the international search 21 DECEMBER 1998		Date of mailing of the international search report 31 DEC 1998
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 308-7724		Authorized officer / PHAN T. H. PALMER / <i>PHAN T. H. PALMER</i> Telephone No. (703) 308-4848

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US98/20081

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

S (FIBER TIP# OR FIBRE TIP#) (P) (METAL LAYER# OR METAL COATING);
S NITRIC OXIDE; S (PROTEIN OR PEPTIDE); S DYE-LABELED;
S NITRIC-OXIDE BINDING COMPOUND